

Callus Culture

D.R.Awad,
Dept. of Botany,
RSM Latur
12.08.2021

HISTORY

- 1924-Callus culture of carrots (*Daucus carota*)- R. Blumenthal and P. Meyer- Pathological implications, compared callus tumour growth. 1927-L. Rehwal- Cultivation of callus from carrot slices. P. Boysen-Jensen growth promoting substances found in plant shoot tip would diffuse across a wound covered with gelatin. 1928
- Frits Went collected this growth substance from coleoptile tip in tiny blocks of Agar.

- 1934 F. Kogl, A. J. Hagen Smith and H. Erxleben isolation and chemical analyses of the substance- plant hormone or auxin- indole-3-acetic acid (IAA) 1937-R. Gautheret and differentiated carrot tissues 1939
- R. J. Gautheret and Nobecourt, France, growth of callus from carrot cambium when using auxin in the nutrient medium. P. R. White (USA)
- Excised root tips of potatoes (*Lycopersicon*) in continuous culture. 1939
- White reported successful culture of tobacco (*Nicotiana*) callus.

- 1941 J. van Overbeek, M. E. Conklin and Albert F. Blakeslee- Coconut milk stimulated callus formation in cultures of excised embryos of jimson weed (*Datura stramonium*). 1943
- White A Handbook of Plant Tissue Culture, accumulated knowledge of PTC.

CALLUS CULTURE

- Callus : An unorganised mass of loosely arranged parenchymatous cells which develop from parent tissues due to proliferation of cells.
- Angiosperms , gymnosperms , pteridophytes and bryophytes.
- Has the potentiality to produce normal roots and embryoids-plantlets
- Callus culture: development of an unorganised mass of cells from an explant on an artificial medium supplemented with suitable PGR when provided with appropriate environment (i.e. Proper incubation conditions)

Physical Appearance Of A Callus

- **HARDNESS** : Hard (due to lignification of cell walls), brittle or sometimes soft
- **COLOUR** : Dirty or off white , creamish to brown or light green to dark green. Degree of darker pattern varies from plant to plant and mainly depends upon the quantity of polyphenols present in the plant species. Higher the polyphenol content darker the callus appear, i.e. Brown coloration on the culture.

Development of Aseptic Callus Cultures

- EXPLANTS : Juvenile tissues, seedlings , young shoots, buds , root tips, developing embryos: fruits, floral parts, tubers and bulbs.
- CULTURE MEDIUM :
- Media: MS
- Hormonal balance: Auxin: Cytokinin= 1
- Carbon source :sucrose (3% w/v)
- ph = 5.6 -6.0, optimum 5.8
- Gelling agent

- **Surface Sterilization**
- Wash throughly tween-20 under running tap --
- Antifungal (Bavistin:0.04%) water Antibacterial agents(Streptomycin sulfate;0.04 %)_UNDER_ROOM
- ↓
- 15-20 min Double Distilled Water
- ↓
- washing 70 % Ethanol treatment
- ↓
- 1 min / dip Under Laminar Air Flow HgCl₂ treatment(0.04 or 0.02 %)
- ↓
- 4-5 min Washing with Autoclaved Water

Inoculation and Incubation

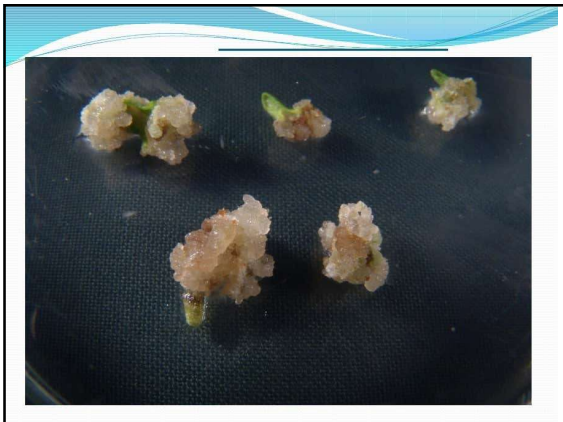
- **Nodal explant**
- Cutting of the nodal end that comes in contact with the surface sterilant with the sterile surgical blade
- Place the explant in vertical position on medium supplemented with appropriate PGR with the help of forceps.
- **Leaf segments, buds, root tip etc.**
- Segmentation of the explant into 2-3 parts i.e. Basal medium and tip with sterile surgical blade
- Place the explant with their abaxial surface in contact with the medium with the help of sterile inoculating forceps.



Incubation :

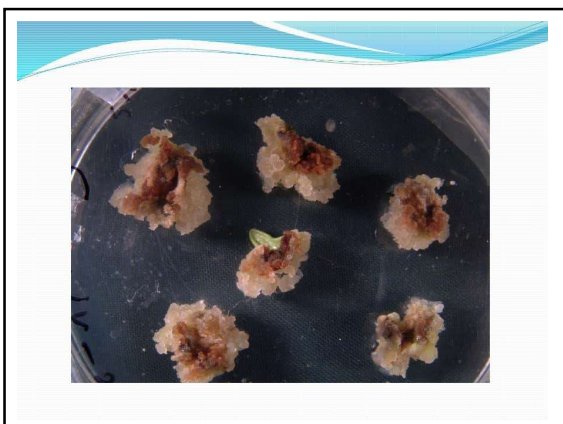
- Incubation conditions : The Environment Temperature : 25 ± 2°C
- Light : 5000-10,000 luxm Duration of incubation: 16 hr light 08 hr dark

- **How a piece of explant gets converted into the callus ?**
- Produced from the outer layers of cortical cells in a stem explant by repetitive division of cells. These dividing cells create pressure on the epidermis --- rupturing of the epidermis exposing newly formed callus Separation of callus and then sub culturing it. 2-3 weeks to grow (10-15 days) but sometimes 4 weeks.



• **SUBCULTURING OF THE CALLUS CULTURE**

- 3-4 weeks
 - 250-500mg approx. pieces -----transferred to the fresh media. NEED:
 - 1) Nutrition depletion
 - 2) Accumulation of toxic substances
 - 3) Drying of media
- ADVANTAGES :1) Maintains the state of viability of cells 2) Provides fresh instalment of media for further growth



• **DISADVANTAGES**

- 1) Cells lose the power to regenerate to a plantlet.
- 2) Chromosomal aberrations: polyploidy and aneuploidy. Polyploid cells appear to originate through endoreduplication (additional rounds of DNA replication without intervening cell division) ; aneuploid cells- anaphase irregularities
- 3) Non chromosomal changes : changes in metabolic pathways and alteration in composition of media.
- 4) Selection of explant size highly dependent upon the type of glassware is being used for culture purpose.

• **Flow sheet (sub culturing process)**

- Callus → work station (glass plate, sterile) cutting of the callus into small pieces (250-500 mg approx) fresh media having appropriate composition and hormonal balance OR Agitation can be done (25-150 rpm) ----- fresh media